

Review Article



Roles of Probiotics in Farm Animals: A Review

Ahmadreza Mirzaei^{1,*} , Seyed Amin Razavi² , Daryoush Babazadeh² , Richard Laven³ , and Muhammad Saeed⁴

¹ Post-Doctoral fellow at College of Veterinary Medicine, University of Florida, Florida, USA

² Department of Clinical Sciences, School of Veterinary Medicine, Shiraz University, Shiraz, Iran

³ School of Veterinary Science, Massey University, Private Bag, Palmerston North, New Zealand

⁴ Department of Animal Nutrition, College of Animal Sciences and Technology, Northwest A&F University, Yangling, China

* **Corresponding author:** Ahmadreza Mirzaei, College of Veterinary Medicine, University of Florida, Florida, USA. Email: ahmadrez.mirzaei@ufl.edu

ARTICLE INFO

Article History:

Received: 10/06/2022

Accepted: 15/07/2022



Keywords:

Antibiotic

Diet

Growth performance

Milk yield

Poultry

ABSTRACT

There are many reports of the positive effects of probiotics on gastrointestinal tract (GIT) microorganisms and the immunological systems of their hosts. Probiotics have prophylactic and metaphylactic properties. The two main mechanisms of action of probiotics seem to be the release of compounds with beneficial effects and direct interaction with the cells of the host. The aim of this review was to evaluate the benefits of probiotic use in farm animals and to identify how they influence farm animal performance. The published data suggest that dietary supplementation of probiotics can improve the growth performance, nutrient digestibility, and immune response of farm animals, including cows, sheep, goats, pigs, aquacultures, and poultry. In ruminants, studies have shown that probiotics can significantly enhance the immune response, milk yield, food digestibility, and weight gain, particularly in ruminants exposed to stressful conditions. This is also the case in aquaculture as probiotics have been shown to enhance growth and reproduction traits, provide protection against pathogens, have positive effects on immunity, optimize digestion, and increase water quality. In horses, there is still controversy about the advantages of probiotic supplementation. In addition, some studies showed valuable effects of using probiotics on treatments of GIT diseases, and some studies showed adverse effects of supplementation of probiotics in horses. In poultry, balancing the intestinal microflora is not achievable but controlling the population of microflora is possible and studies showed that supplementation of probiotics could influence some aspects such as improving performance and health parameters. So, probiotics are used by almost all farmers who are trying to reduce antibiotic resistance. In conclusion, dietary supplementation of probiotics to farm animals has many notable influences on their performance, immune system, and diet digestion.

1. Introduction

Probiotics have attracted great attention due to their positive effects on gut microbiota and immunological systems in humans and livestock^{1,2}. Probiotics are used as prophylaxis and for therapeutic purposes in both humans and farm animals^{3,4}, with probiotics being a safe and viable alternative to the use of antibiotics growth promoters which can result in antibiotic resistance in microbial population⁵. When used in farm animals instead of antibiotics dietary supplementation of probiotics can

improve growth performance, nutrient digestibility, and immune response and reduce the incidence of diseases^{1,5}. In addition, probiotics increase the oxygen concentration in the gastrointestinal tract (GIT) in a multiplicity of ways including through depleting oxygen-scavenging compounds, such as nitrates. Moreover, probiotics secrete hydrolytic enzymes that can hydrolyze bacterial toxins and inactivate toxin receptors, decreasing toxin-mediated infections in farm animals⁶.

Probiotics decrease pathogenic microorganisms in the GIT and establish a better microbial population in the GIT⁷. Probiotics can also activate an immunologic response and assemble a series of immunoregulatory molecules⁸. Some probiotics produce metabolites that can modulate metabolic pathways in the cell, such as bacteriocins, amines, and hydrogen peroxide. These metabolites can adjust cell apoptosis, proliferation, inflammation, and differentiation⁹.

Microorganisms used as probiotics should be nonpathogenic, optimize host health, and improve GIT function. The microorganisms commonly used as probiotics in animal diets are principally bacteria, usually Gram-positive bacteria. *Lactobacillus* and *Enterococcus* are natural flora in the alimentary tract of most animals and are usually present in amounts of 10^7 - 10^8 and 10^5 - 10^6 CFU/g, respectively¹⁰. *Lactobacillus acidophilus* (*L. acidophilus*), *L. lactis*, *L. plantarum*, *L. bulgaricus*, *L. casei*, *L. helveticus*, *L. salivarius*, *Bifidobacterium* spp., *Enterococcus faecium* (*Ent. faecium*), *Ent. faecalis*, *Streptococcus thermophilus*, *Escherichia coli* are the most commonly used probiotic bacteria in farm animals and probiotic fungi such as *Saccharomyces cerevisiae* (*Sacc. cerevisiae*) and *Sacc. boulardii* are the most commonly used probiotic fungi¹¹⁻¹⁵. Of note, no other substances should be supplemented during the administration of probiotics such as antibiotics, disinfectants, and materials that interrupt with the mechanism of action of probiotics. In addition, water used for dilution should not contain chlorine or disinfectants¹⁰. Factors, such as the optimal selection of microbial strains, host species, and age are critical in the selection of probiotics¹⁵. Given the positive effects of dietary supplementation of probiotics on farm animals' immune response and growth performance, the current study aimed to review the benefits of probiotics and their mechanism of action in farm animals.

2. Usage and benefits

The microbial balance of the GIT in farm animals is critical as it can affect feed efficiency, digestive welfare, and health in farm animals¹⁰. It is influenced by many factors including diet ingredients, farm management, feeding practices, and productivity limitations, for example, a change of diet from forage to a high-concentrate diet results in a change in the microbial population of the rumen and can cause ruminal acidosis. Weaning is a critical period for the gut microbiota as the change in the diet results in major changes in that microbiota, which if not properly managed increases the susceptibility of young animals to pathogen colonization¹⁶.

Animal products are closely related to consumers' health. Animal intestinal pathogens, such as *Campylobacter*, *Salmonella*, *Listeria*, and *Yersinia*, contaminate foods and result in zoonosis disease, outbreaks of which threaten public health and bring about economic loss¹⁷. Thus, producers need to provide animal breeding methods to increase meat quality and safety, pay attention to animal welfare, and respect the natural environment.

According to some estimations, the world population is growing very fast and could reach 10 billion in 2050. Therefore, it could cause food security challenges worldwide¹⁸. Antibiotics have been used commonly to change the alimentary tract microorganism and increase productivity and animal growth¹⁹. Prolonged use of antibiotics results in the development of drug-resistant microbiota. Drug-resistant development in microbiota threatens consumers' health, causes a negative impact on the environment, and increases foodborne allergies²⁰⁻²². To confront these challenges, probiotics could be beneficial. Therefore, the use of probiotics as dietary supplementation has received great concerted attention.

3. Mechanism of action

The metabolic activities, survivability of the probiotic throughout the gut, and the used strains are essential for the optimal efficacy of probiotics²³. The combination of several mechanisms has indicated the positive effects of probiotics on animals' health. Two main mechanisms of action of probiotics include the release of compounds with inhibitory effects and direct interaction between cells²⁴⁻²⁶. Notably, organic acids, such as lactic or acetic acid, produced by bacterial probiotics can reduce the GIT pH, making it less favorable for pathogen colonization, particularly in monogastric animals²⁷. Some probiotics release antimicrobial compounds, such as peptides (bacteriocins), hydrogen peroxide, and biosurfactants that prevent the pathogenic bacteria growth or release enzymes hydrolyzing bacterial toxins²⁸. Probiotics have a higher affinity to adhesion sites and compete with GIT pathogens for nutrients^{29,30}. In addition, studies have revealed that probiotics positively affect immune response³¹; for instance, administration of *L. acidophilus*, *L. salivarius*, and *L. plantarum* at the dosage of 10^7 - 10^8 CFU/g can reduce the occurrence of diarrhea in calves³².

Moreover, dietary probiotic supplementation increases disease resistance and decreases metabolic stress and mortality²⁷. Some probiotics play an influential role in detoxifying and metabolizing inhibitory chemicals, such as nitrates and amines or oxygen scavenging²³, which is of utmost importance for GIT anaerobic ecosystems. The mechanism of probiotics action in human GIT is almost similar to that of animals and has health and nutrition benefits¹⁰.

4. Probiotics in farm animal

4.1. Probiotics in ruminant

Most bacteria in the ruminant intestinal tract belong to the phyla *Bacillota* (*Firmicutes*) and *Bacteroidota* (*Bacteroidetes*) with species in the phyla *Actinobacteria*, *Proteobacteria*, and *Verrucomicrobia* also being common. For example, the microbiological ecosystem of the rumen in the ruminant is complex, yet nearly 99% of bovine rumen bacteria and 96% of ovine rumen bacteria belong are *Firmicutes* or *Bacteroidetes*¹⁵. Nevertheless, the

composition of the rumen microbiota is sensitive to change based on feed and animal status. So, the key times for which probiotics have been shown to be especially beneficial are at weaning, after calving, and after dietary changes from forage to high-concentrate diets for the gut microbiota. Probiotics, such as *Sacc. cerevisiae*, *Lactobacilli*, *Aspergillus oryzae*, *Bacillus*, and *Enterococcus*, are commonly at such time points and positively affect the ruminant health^{12,14,23,33-37}. Some studies have indicated that milk yield increases in cows fed a diet supplemented with probiotics. In addition, some probiotics can enrich milk secretion, such as *Bacillus subtilis*, *Sacc. cerevisiae*, and *Ent. faecalis*³⁸. *Bifidobacterium bifidum* also prevents milk allergy reactions in dairy cows³⁹. According to a previous study⁴⁰, probiotics containing *Ent. faecium* and *Sacc. cerevisiae* increases milk yield by 2.3 liters per dairy cattle daily. Another study showed that active yeast probiotics could enhance milk yield by nearly 1.2 g/kg of body weight in dairy cows, and increase dry matter intake in the ruminant by about 0.44 g/kg of body weight, but had no effects on milk protein percentage⁴¹. A study on goats revealed that the probiotic combination of *L. reuteri* DDL 19, *L. alimentarius* DDL 48, *Ent. faecium* DDE 39, and *Bifidobacterium bifidum* DDBA obtained from healthy goats could enhance the body weight of goats fed the combination for two months by 9%⁴². Some probiotics, such as *B. subtilis* and *B. amyloliquefaciens* can upgrade intestinal maturation by stimulating GH/IGF-1 hormone⁴³. Some studies have indicated that probiotics can enhance the feed digestibility and immune system in ruminants¹⁶⁻¹⁸. A previous study revealed that probiotics (a mixture of *L. acidophilus* NP51 and *Propionibacterium freudenreichii* NP24) could increase the digestion of neutral detergent fiber, crude protein, and milk production by nearly 7.6% in ruminants⁴⁴. In juvenile calves, mixed probiotics of *L. acidophilus*, *L. salivarius*, and *L. plantarum* at a dosage of 10⁷-10⁸ CFU/g reduced diarrhea occurrence⁴⁵. *Lactococcus lactis* produces an antimicrobial peptide named nisin, and infusion of nisin in the mammary gland in dairy cows can treat mastitis caused by *S. aureus*⁴⁶. Teat spray containing *Lactobacillus* spp. has a positive effect on mammary gland condition and improves the functions of the teat sphincter⁴⁷. Adding probiotics to young stressed calves' diet can improve immunity response and alleviate acidosis in cows with rumen acidosis⁴⁸. In addition, probiotics release antimicrobial components that can reduce zoonotic pathogens and limit the risk of foodborne diseases in humans¹⁰. Probiotics can control ammonia production in the rumen¹³ and mitigate methane excretion, which reduces ruminants' negative impact on the environment as methane produced by ruminants is responsible for 3-5% of global warming¹⁷. Probiotics containing *Rhodopseudomonas palustris* improve the growth performance of rumen microorganisms, increase microbial fermentation and optimize rumen microbial balance^{49,50}.

4.2. Probiotics in non-ruminant

Bacterial strains that are commonly used in probiotics in monogastric animals are *Lactobacillus* spp., *Enterococcus*

spp., *Pediococcus* spp., *Bacillus* spp., and yeasts, such as *Sacc. boulardii* that target cecum and colon¹⁷.

The large intestine of horses is home to different microorganisms composed of bacteria, protozoa, and fungi that significantly impacting horse health and performance^{51,52}. In equine medicine, some diseases such as acute colitis, equine grass sickness, and laminitis have all been linked to changes in the equine intestinal microbiota^{52,53}. The most common phylum that exists in equine feces is *Firmicutes* which accounts for 46-70% of all identified bacteria following *Bacteroidetes*, *Proteobacteria*, *Verrucomicrobia*, *Actinobacteria*, and *Spirochaetes* each accounting for 0-15%^{52,54,55}. Compared to horses with diarrhea, healthy horses have higher levels of *Actinobacteria* and *Spirochetes* and higher levels of *Fusobacteria*^{52,56}. The large colon is the site of several diseases in equines and probiotics are intended to target this site. Species *Lactobacillus*, *Bifidobacterium*, and *Enterococci* which are commonly used in probiotics are not the most abundant species in the horse's large intestine^{52,55}. Probiotics can modulate the immune system by strengthening the intestinal barrier, promoting the survival and growth of epithelial cells, and inducing the production of IgA and β -defensin, which inhibits pathogen growth⁵⁷. Horses receiving *Sacc. boulardii* had a shorter period of watery diarrhea, but this finding should be interpreted with caution since only seven animals were used per group, and there was no difference between treatment groups⁵⁸. Furthermore, obtained results may be confounded by treatment protocols. In another study, *Sacc. boulardii* was administered as an adjunctive therapy to horses with antimicrobial-associated enterocolitis, and no differences were observed in the treatment groups⁵⁹. Another study assessed *L. pentosus* in a randomized placebo-controlled clinical trial, and its use was associated with a higher incidence of diarrhea, fever, and anorexia⁶⁰. Two other studies report similar findings that the use of probiotics increased the risk of diarrhea in neonatal foals^{61,62}. In contrast, A study that used multistrain equine-derived probiotics reported an increase in weight gain and a decrease in the incidence of diarrhea in neonatal foals⁶³. Two studies on horses tried to determine the effect of probiotics on Salmonella shedding, but the results were not promising^{64,65}.

Various hypotheses have been mentioned as the reason why probiotics do not work effectively in horses, compared to other animals, especially ruminants. Most probiotics supplements in horses fail to meet the FAO and WHO (2002)⁵⁶ definitions because of quality control issues and a lack of specific research currently existing. Several studies indicated that some probiotics in the market that are used as a supplementation in horses did not contain the amounts specified on the product label and any quantity of some microorganisms^{62,66,67}.

Another hypothesis related to the lack of effectiveness of probiotics in horses is that physiological decreases in gastric pH protect horses from potential pathogenic microorganisms, but this reduction of gastric pH can also negatively impact the viability of microorganisms in equine

probiotics⁶⁸. Another disadvantage of using probiotics in horses is that many products are based on human studies data that may not apply to horses. Furthermore, several studies have shown that antimicrobial-resistant genes could transfer by probiotics in horses⁶⁷, which needs further investigation. Thus, more studies are required to capitalize on probiotics' potential benefits in equines.

Supplementation of probiotics could reduce diarrhea prevalence in piglets and prevent the colonization of pathogenic organisms, such as *E. coli*, *Clostridium difficile*, *Clostridium perfringens*, *Salmonella*, and *Listeria*, parasites (*Isospora* and *Cryptosporidium*) or viruses (*Coronavirus* and *Rotavirus*)⁶⁹. These pathogenic organisms can result in growth reduction and diarrhea in piglets¹⁷. Weaning is a critical moment in pig production when animals are exposed to the stressful condition such as nutrition changes from milk to a vegetable base diet, and animal transfer to another farm cause stress and consequently disturb immunological function and harms pigs' GIT. Studies reported that probiotics positively influence the immune system in stress conditions in pigs notably during weaning^{69,70}. Probiotic supplementation can increase dry matter intake and average live weight in pigs⁷¹.

4.3. Probiotics in aquaculture

FAO (2020) reported that 179 million tons of fish were produced worldwide in 2018, which cost about 400 billion USD. Antibiotics are commonly used in the aquaculture industry to meet the increasing demand for fish meat. However, antibiotics increase the drug-resistant bacteria transmitted by food to humans^{72,73}. The probiotics supplementation in water-dwelling animals enhances growth and reproduction, provides protection against pathogens, exerts positive effects on immunity, optimizes digestion, increases water quality, and can be an alternative to antibiotics⁷⁴. One of the probiotics commonly used in fish farms is *Bacillus subtilis*, which can prevent harmful pathogens, such as *Vibrio*, *Pseudomonas*, *Aeromonas*, *Clostridium*, *Streptococcus*, *Flavobacterium*, *Acinetobacter*, and white spot syndrome virus in fishes⁷⁵. Some bacterial strains, such as *L. lactis*⁷⁶ and *L. plantarum* VSG-3⁷⁷ are commonly used in fish farms. Gram-negative facultatively anaerobic bacteria are dominant in the GIT of aquaculture.

According to some studies, microalgae, such as *Dunaliella tertiolecta*, *Dunaliella salina*, *Isochrysis galbana*, *Phaeodactylum tricornutum*, and *Tetraselmis suecica*, increase the survival rate of aquatic animals^{78,79}. A few studies reported the positive effects of *Sacc. cerevisiae* (yeast) on aquatic animals^{80,81}. Farmers supplement probiotics to water circulation or to the fish diet⁸². Some probiotics are immunostimulants in aquatic animals that could be a single or combination of bacterial strains^{83,84}. Proper timing and dosage are essential for obtaining the expected outcomes in fish farms¹⁸. Probiotics could optimize the health conditions of aquatic animals. Some studies reported that probiotics

containing *B. pumilus*⁸⁵ and *L. plantarum*⁸⁶ can improve the health conditions of Nile tilapia and decrease the fatality rate caused by Lake Virus infection in Nile tilapia⁸⁷. Probiotics containing *Pseudomonas* I-2 can decrease diseases caused by *vibrio* microorganisms in fishes⁸⁸. Probiotics with *Pediococcus acidilactici* can prevent vibriosis in white-leg shrimp⁸⁹. Dietary supplementation of probiotics plus *Lactobacillus*, *Saccharomyces*, *Bacillus*, *Bifidobacterium*, and Photosynthetic bacteria can increase the growth rate and larval metamorphosis in white-leg shrimp larvae. These strains also decrease the number of *vibrio* pathogens and encumber incomplete molting during the development in White leg shrimp larvae⁹⁰. Some studies showed that probiotics containing *Pediococcus pentosaceus* and *S. hemolyticus* could reduce the prevalence of white spot syndrome virus in white leg prawns⁹¹. The *Sacc. cerevisiae* can be substituted as live food in the cultivation of some fish species, such as clownfish, Catla, hybrid striped bass, Japanese flounder, and Nile tilapia^{18,92-95}. Probiotics have a positive effect on immune responses in fish. For instance, probiotics, including *Bacillus* activates, the humoral and cell-mediated immunologic response in fish^{96,97}. In addition, probiotics can increase digestive enzyme processes, growth performance, and feed intake in aquaculture. Probiotics produce enzymes that improve nutrient digestion in aquacultures, such as protease, carbohydrase, and lipase^{98,99}. Some studies showed that probiotics could enhance water quality by nourishing water properties, preventing diseases, and optimizing fish habitats, also reducing metabolic wastes^{100,102}. Moreover, probiotics can increase water quality by decreasing pathogen microbes, nitrogen, and phosphate contamination in the sediments¹⁰³⁻¹⁰⁵.

4.4. Probiotics in poultry

The global concern about antibiotic residue in poultry products is increasing, and it is clear that more attempts should be conducted to find the best antibiotic alternative in poultry diets. One of the best approaches to control poultry diseases, perform prophylactic treatment, and recover the birds after antibiotic treatment is definitely probiotic bacteria¹⁰⁶.

However, balancing the intestinal microflora is not achievable but controlling the population of microflora is possible to occur. The administration of the beneficial bacteria is possible via different methods in the poultry industry¹⁰⁷⁻¹⁰⁹. Currently, *in ovo* administration of probiotics is considered in the poultry production system. This method of administration aimed to stimulate the immune system, control intestinal microflora, and improve the characteristics of gut health in the first days of life. *In ovo* feeding and vaccination are studied along with the administration of probiotics in hatchery¹⁰⁹.

The hatched chickens are exposed to many different bacteria in the first days of growing. Oral administration of probiotics or even prebiotics can improve performance and health parameters¹⁰⁶⁻¹⁰⁹. The probiotic agents usually do

not absorb from the intestines and have no side effects on internal organs. However, some of these agents, such as *Sacc. cerevisiae*, can reduce the adverse side effects of aflatoxins on the liver^{109,110}. The most common agents of synthetic probiotics in poultry industries are *Lactobacillus* spp., *Bifidobacterium bifidum*, *B. subtilis*, *S. salivarius* sub spp. *Thermophilus*, *Aspergillus oryzae*, and *Sacc. cerevisiae*^{106,109-110}. The positive effects of some natural sources of probiotics and phytobiotics, such as Kefir, are also reported^{111,112}. The most important challenge of probiotic administration in different growing ages is administration duration. There are many commercial recommendations for using industrial products in a limited time to improve poultry performance parameters. However, it seems necessary to design some structured protocol studies on supplementing probiotics in different ages and health conditions.

5. Risks of using probiotic

Safety evaluation of probiotic strains is essential because probiotics might lead to adverse effects, such as transferring virulence factors and antimicrobial resistance, hemolytic potential, and production of toxic biochemicals¹¹³. Moreover, the potential risk of transmission of antimicrobial-resistant genes in the probiotic strain to other bacteria of the normal flora of GIT is more critical. Rarely, infectious diseases such as endocarditis, bacteremia, pneumonia, meningitis, and septic arthritis caused by *Lactobacillus* and *Enterococcus* are reported in immuno-compromised patients¹¹⁴. Some authors discourage using *Enterococcus* as a probiotic because *Enterococcus* is more likely to develop vancomycin-resistant genes and can transfer to neighboring pathogens and enhance the virulence of those pathogens⁶. Discovering the origin of GIT microbiota is necessary for ascertaining the source of a microorganism in the GIT. In addition, the possibility of probiotic supplementation that could contaminate the human food chain must be considered.

6. Conclusion

In conclusion, farmers use additives to increase growth performance due to the increasing demand for meat production. Some additives, such as antibiotics, are growth stimulators but have adverse effects on human and animal health, such as increasing antimicrobial resistance. Alternatives like probiotics could be a safer substitution. Probiotics are used as prophylaxes and for therapeutic purposes in humans and farm animals. In addition, probiotics positively affect the immune system, particularly in stressful conditions, increase performance such as growth performance and milk yield, enhance nutrient digestibility, and prevent some diseases in farm animals. More studies are needed to evaluate the effects of probiotics on farm animal performance and the safety of these additives.

Declarations

Competing interests

There is no conflict of interest.

Authors' contribution

The final manuscript draft was reviewed by all authors, who also gave their approval.

Funding

No funding.

Ethical considerations

Ethical issues (including plagiarism, consent to publish, misconduct, data fabrication and/or falsification, double publication and/or submission, and redundancy) have been checked by all the authors.

Acknowledgments

The authors wish to acknowledge everyone who helped during the writing of this manuscript.

References

1. Celiberto LS, Bedani R, Rossi EA, and Cavallini DC. Probiotics: The scientific evidence in the context of inflammatory bowel disease. *Crit Rev Food Sci Nutr*. 2017; 57(9): 1759-1768. DOI: <https://doi.org/10.1080/10408398.2014.941457>
2. Chen J, Wang Q, Liu CM, and Gong J. Issues deserve attention in encapsulating probiotics: Critical review of existing literature. *Crit Rev Food Sci Nutr*. 2017; 57(6): 1228-1238. Doi: <https://doi.org/10.1080/10408398.2014.977991>
3. de Llano DG, Gil-sánchez I, Esteban-fernández A, Ramos AM, Fernández-díaz M, Cueva C, et al. Reciprocal beneficial effects between wine polyphenols and probiotics: An exploratory study. *Eur Food Res Technol*. 2016; 243: 531-538. Available at: https://www.infona.pl/resource/bwmeta1.element.springer-doi-10_1007-S00217-016-2770-5
4. Abushelaibi A, Al-mahadin S, El-tarabily K, Shah NP, and Ayyash M. Characterization of potential probiotic lactic acid bacteria isolated from camel milk. *LWT Food Sci Technol*. 2017; 79: 316-325. DOI: <https://doi.org/10.1016/j.lwt.2017.01.041>
5. Srinivas B, Rani GS, Kumar BK, Chandrasekhar B, Krishna KV, Devi TA, et al. Evaluating the probiotic and therapeutic potentials of *Saccharomyces cerevisiae* strain (OBS2) isolated from fermented nectar of toddy palm. *AMB Express*. 2017; 7: 2. DOI: <https://doi.org/10.1186%2Fs13568-016-0301-1>
6. Alayande KA, Aiyegoro OA, Ateba CN. Probiotics in animal husbandry: Applicability and associated risk factors. *Sustainability*. 2020; 12: 1087. Doi: <https://doi.org/10.3390/su12031087>
7. Mountzouris KC, Balaskas C, Xanthakos I, Tzivinikou A, and Fegeros K (2009). Effects of a multi-species probiotic on biomarkers of competitive exclusion efficacy in broilers challenged with *Salmonella enteritidis*. *Bri Poult Sci*. 50(4): 467-478. DOI: <https://doi.org/10.1080/00071660903110935>
8. Corcionivoschi N, Drinceanu D, Pop IM, Stack D, Ștef L, Julean C, et al. (2010). The Effect of probiotics on animal health, review. scientific papers: Animal Science and Biotechnologies. 43(1): 35-41.
9. Plaza-Díaz J, Ruiz-Ojeda FJ, Gil-Campos M, and Gil A. Mechanisms of action of probiotics. *Adv Nutr*. 2019; 10(1): S49-S66. DOI: <https://doi.org/10.1093/advances/nmy063>

10. Chaucheyras-Durand F and Durand H. Probiotics in animal nutrition and health. *Benef Microbes*. 2010; 1(1): 3-9. DOI: <https://doi.org/10.3920/bm2008.1002>
11. McFarland LV. Meta-analysis of probiotics for the prevention of antibiotic associated diarrhea and the treatment of *Clostridium difficile* disease. *Am J Gastroenterol*. 2006; 101(4): 812-822. DOI: <https://doi.org/10.1111/j.1572-0241.2006.00465.x>
12. Hossain M, Ko S, Kim G, Firman J, and Yang C. Evaluation of probiotic strains for development of fermented *Alisma canaliculatum* and their effects on broiler chickens. *Poult Sci*. 2012; 91(12): 3121-3131. DOI: <https://doi.org/10.3382/ps.2012-02333>
13. Razavi SA, Pourjafar M, Hajimohammadi A, Valizadeh R, Naserian AA, Laven R, et al. Effects of dietary supplementation of bentonite and *Saccharomyces cerevisiae* cell wall on acute-phase protein and liver function in high-producing dairy cows during transition period. *Trop Anim Health Prod*. 2019a; 51: 1225-1237. DOI: <https://doi.org/10.1007/s11250-019-01815-3>
14. Razavi SA, Pourjafar M, Hajimohammadi A, Valizadeh R, Naserian AA, Laven R, et al. Effects of dietary supplementation of bentonite and yeast cell wall on serum blood urea nitrogen, triglyceride, alkaline phosphatase, and calcium in high-producing dairy cattle during the transition period. *Comp Clin Pathol*. 2019b; 28: 419-425. DOI: <https://doi.org/10.1007/s00580-018-2849-4>
15. Mirzaei A, Hajimohammadi A, Badieli K, Pourjafar M, Naserian AA, and Razavi SA. Effect of dietary supplementation of bentonite and yeast cell wall on serum endotoxin, inflammatory parameters, serum and milk aflatoxin in high-producing dairy cows during the transition period. *Comp Clin Pathol*. 2020; 29: 433-440. DOI: <https://doi.org/10.1007/s00580-019-03074-y>
16. Al-Shawi SG, Dang DS, Yousif AY, Al-Younis ZK, Najm TA, and Matarneh SK. The potential use of probiotics to improve animal health, efficiency, and meat quality: A review. *Agriculture*. 2020; 10(10): 452. DOI: <https://doi.org/10.3390/agriculture10100452>
17. Markowiak P and Śliżewska K. The role of probiotics, prebiotics and synbiotics in animal nutrition. *Gut Pathog*. 2018; 10: 21. DOI: <https://doi.org/10.1186/s13099-018-0250-0>
18. Anee IJ, Alam S, Begum RA, Md Shahjahan R, and Khandaker AM. The role of probiotics on animal health and nutrition. *J Basic Appl Zool*. 2021; 82: 52. DOI: <https://doi.org/10.1186/s41936-021-00250-x>
19. Deng Y, Xu L, Liu S, Wang Q, Guo Z, Chen C, et al. What drives changes in the virulence and antibiotic resistance of *Vibrio harveyi* in the South China Sea? *J Fish Dis*. 2020a; 43(8): 853-862. DOI: <https://doi.org/10.1111/jfd.13197>
20. Li XY, Duan YL, Yang X, and Yang XJ. Effects of *Bacillus subtilis* and antibiotic growth promoters on the growth performance, intestinal function and gut microbiota of pullets from 0 to 6 weeks. *Animal*. 2020; 14(8): 1619-1628. DOI: <https://doi.org/10.1017/S1751731120000191>
21. Deng Z, Luo XM, Liu J, and Wang H. Quorum sensing, biofilm, and intestinal mucosal barrier: Involvement the role of probiotic. *Front Cell Infect Microbiol*. 2020b; 10: 538077. DOI: <https://doi.org/10.3389/fcimb.2020.538077>
22. Han T, Zhang Q, Liu N, Wang J, Li Y, Huang X, et al. Changes in antibiotic resistance of *Escherichia coli* during the broiler feeding cycle. *Poult Sci*. 2020; 99(12): 6983-6989. DOI: <https://doi.org/10.1016/j.psj.2020.06.068>
23. Chaucheyras-Durand F, Walker ND, and Bach A. Effects of active dry yeasts on the rumen microbial ecosystem: Past, present and future. *Anim Feed Sci Technol*. 2008; 145(1-4): 5-26. DOI: <https://doi.org/10.1016/j.anifeedsci.2007.04.019>
24. Parvez S, Malik KA, Ah Kang S, and Kim HY. Probiotics and their fermented food products are beneficial for health. *J Appl Microbiol*. 2006; 100(6): 1171-1185. DOI: <https://doi.org/10.1111/j.1365-2672.2006.02963.x>
25. Boirivant M and Strober W. The mechanism of action of probiotics. *Curr Opin Gastroenterol*. 2007; 23: 679-692. DOI: <https://doi.org/10.1097/mog.0b013e3282f0cfcf>
26. Musa HH, Wu S, Zhu C, Seri H, and Zhu G. The potential benefits of probiotics in animal production and health. *J Anim Vet Adv*. 2009; 8(2): 313-321. Available at: <https://www.medwelljournals.com/abstract/?doi=javaa.2009.313.321>
27. Servin AL. Antagonistic activities of lactobacilli and bifidobacteria against microbial pathogens. *FEMS Microbiol Rev*. 2004; 28(4): 405-440. DOI: <https://doi.org/10.1016/j.femsre.2004.01.003>
28. Buts JP. Exemple d'un médicament probiotique. In: Rambaud JC, Buts JP, Corthier G, Flourié B, editors. *Saccharomyces boulardii lyophilisé. Flore microbienne intestinale*. Montrouge, France: John Libbey Eurotext; 2004. p. 221-244.
29. La Ragione RM, Narbad A, Gasson MJ, and Woodward MJ. Competitive exclusion by *Bacillus subtilis* spores of *Salmonella enterica* serotype enteritidis and *Clostridium perfringens* in young chickens. *Vet Microbiol*. 2003; 94(3): 245-256. DOI: [https://doi.org/10.1016/s0378-1135\(03\)00077-4](https://doi.org/10.1016/s0378-1135(03)00077-4)
30. La Ragione RM, Narbad A, Gasson MJ, and Woodward MJ. *In vivo* characterization of *Lactobacillus johnsonii* FI9785 for use as a defined competitive exclusion agent against bacterial pathogens in poultry. *Lett Appl Microbiol*. 2004; 38(3): 197-205. DOI: <https://doi.org/10.1111/j.1472-765x.2004.01474.x>
31. Delcenserie V, Martel D, Lamoureux M, Amiot J, Boutin Y, and Roy D. Immunomodulatory effects of probiotics in the intestinal tract. *Curr Issues Mol Biol*. 2008; 10(1-2): 37-54. <https://pubmed.ncbi.nlm.nih.gov/18525105/>
32. Johnson-Henry KC, Donato KA, Dhen-Tu G, Gordanpour M, and Sherman PM. *Lactobacillus rhamnosus* strain GG prevents enterohemorrhagic *Escherichia coli* O157:H7-induced changes in epithelial barrier function. *Infect Immun*. 2008; 76(4): 1340-1348. DOI: <https://doi.org/10.1128/iai.00778-07>
33. Elghandour MMY, Tan ZL, Abu Hafa SH, Adegbeye MJ, Greiner R, Ugbogu EA, et al. *Saccharomyces cerevisiae* as a probiotic feed additive to non- and pseudo-ruminant feeding: A review. *J Appl Microbiol*. 2020; 128(3): 658-674. DOI: <https://doi.org/10.1111/jam.14416>
34. Chiquette J, Allison MJ, and Rasmussen MA. *Prevotella bryantii* 25A used as a probiotic in early-lactation dairy cows: Effect on ruminal fermentation characteristics, milk production, and milk composition. *J Dairy Sci*. 2008; 91(9): 3536-3543. Available at: [https://www.journalofdairyscience.org/article/S0022-0302\(08\)71068-3/fulltext](https://www.journalofdairyscience.org/article/S0022-0302(08)71068-3/fulltext)
35. Weimer PJ. Redundancy, resilience, and host specificity of the ruminal microbiota: Implications for engineering improved ruminal fermentations. *Front Microbiol*. 2015; 6: 296. DOI: <https://doi.org/10.3389/fmicb.2015.00296>
36. Uyeno Y, Shigemori S, and Shimamoto T. Effect of probiotics/prebiotics on cattle health and productivity. *Microbes Environ*. 2015; 30(2): 126-132. DOI: <https://doi.org/10.1264/jsme.2ME14176>
37. Jouany JP, Mathieu F, Senaud J, Bohatier J, Bertin G, and Mercier M. Effect of *Saccharomyces cerevisiae* and *Aspergillus oryzae* on the digestion of nitrogen in the rumen of defaunated and refaunated sheep. *Anim Feed Sci Technol*. 1998; 75(1): 1-13. DOI: [https://doi.org/10.1016/S0377-8401\(98\)00194-1](https://doi.org/10.1016/S0377-8401(98)00194-1)
38. Ma ZZ, Cheng YY, Wang SQ, Ge JZ, Shi HP, and Kou JC. Positive effects of dietary supplementation of three probiotics on milk yield, milk composition and intestinal flora in Sannan dairy goats varied in kind of probiotics. *J Anim Physiol Anim Nutr (Berl)*. 2020; 104(1): 44-55. DOI: <https://doi.org/10.1111/jpn.13226>
39. Jing W, Liu Q, and Wang W. *Bifidobacterium bifidum* TMC3115 ameliorates milk protein allergy in by affecting gut microbiota: A randomized double-blind control trial. *J Food Biochem*. 2020; 44(11): e13489. DOI: <https://doi.org/10.1111/jfbc.13489>
40. Nocek JE and Kautz WP. Direct-fed microbial supplementation on ruminal digestion, health, and performance of pre- and postpartum dairy cattle. *J Dairy Sci*. 2006; 89(1): 260-261. DOI: [https://doi.org/10.3168/jds.S0022-0302\(06\)72090-2](https://doi.org/10.3168/jds.S0022-0302(06)72090-2)
41. Desnoyers M, Giger-Reverdin S, Bertin G, Duvaux-Ponter C, and Sauvand D. Meta-analysis of the influence of *Saccharomyces cerevisiae* supplementation on ruminal parameters and milk production of ruminants. *J Dairy Sci*. 2009; 29(4): 1620-1632. DOI: <https://doi.org/10.3168/jds.2008-1414>
42. Apás AL, Dupraz J, Ross R, González SN, and Arena ME. Probiotic administration effect on fecal mutagenicity and microflora in the goat's gut. *J Biosci Bioeng*. 2010; 110(5): 537-540. DOI: <https://doi.org/10.1016/j.jbiosc.2010.06.005>
43. Du R, Jiao S, Dai Y, An J, Lv J, Yan X, et al. Probiotic *Bacillus amyloliquefaciens* C-1 improves growth performance, stimulates GH/IGF-1, and regulates the gut microbiota of growth-retarded beef calves. *Front Microbiol*. 2018; 9: 2006. DOI: <https://doi.org/10.3389/fmicb.2018.02006>
44. Boyd J, West JW, and Bernard JK. Effects of the addition of direct-fed microbials and glycerol to the diet of lactating dairy cows on milk yield and apparent efficiency of yield. *J Dairy Sci*. 2011; 94(9): 4616-4622. DOI: <https://doi.org/10.3168/jds.2010-3984>

45. Signorini ML, Soto LP, Zbrun MV, Sequeira GJ, Rosmini MR, and Frizzo LS. Impact of probiotic administration on the health and fecal microbiota of young calves: A meta-analysis of randomized controlled trials of lactic acid bacteria. *Res Vet Sci*. 2012; 93(1): 250-258. DOI: <https://doi.org/10.1016/j.rvsc.2011.05.001>
46. Cao LT, Wu JQ, Xie F, Hu SH, and Mo Y. Efficacy of nisin in treatment of clinical mastitis in lactating dairy cows. *J Dairy Sci*. 2007; 90(8): 3980-3985. DOI: <https://doi.org/10.3168/jds.2007-0153>
47. Alawneh JI, James AS, Phillips N, Fraser B, Jury K, Soust M, et al. Efficacy of a Lactobacillus-based teat spray on udder health in lactating dairy cows. *Front Vet Sci*. 2020; 7: 584436. DOI: <https://doi.org/10.3389/fvets.2020.584436>
48. Krehbiel C, Rust S, Zhang G, and Gilliland S. Bacterial direct-fed microbials in ruminant diets: Performance response and mode of action. *J Anim Sci*. 2003; 81(14): E120-E132. DOI: https://doi.org/10.2527/2003.8114_suppl_2E120x
49. Chen B, Peng M, Tong W, Zhang Q, and Song Z. The Quorum Quenching Bacterium *Bacillus licheniformis* T-1 protects Zebrafish against *Aeromonas hydrophila* infection. *Probiotics Antimicrob Proteins*. 2020a; 12(1): 160-171. DOI: <https://doi.org/10.1007/s12602-018-9495-7>
50. Chen YY, Wang YL, Wang WK, Zhang ZW, Si XM, Cao ZJ, et al. Beneficial effect of *Rhodopseudomonas palustris* on *in vitro* rumen digestion and fermentation. *Benef Microbes*. 2020b; 11(1): 91-99. DOI: <https://doi.org/10.3920/BM2019.0044>
51. Al Jassim RA, Andrews FM. The bacterial community of the horse gastrointestinal tract and its relation to fermentative acidosis, laminitis, colic, and stomach ulcers. *The Veterinary Clinics of North America*. *Equine Pract* 2009; 25:199-215
52. Costa MC and Weese JS. The equine intestinal microbiome. *Anim Health Res Rev*. 2012; 13(1): 121-128. DOI: <https://doi.org/10.1017/s1466252312000035>
53. Moreau MM, Eades SC, Reinemeyer CR, Fugaro MN, and Onishi JC. Illumina sequencing of the V4 hypervariable region 16S rRNA gene reveals extensive changes in bacterial communities in the cecum following carbohydrate oral infusion and development of early-stage acute laminitis in the horse. *Vet Microbiol*. 2014; 168(2-4): 436-441. DOI: <https://doi.org/10.1016/j.vetmic.2013.11.017>
54. Shepherd ML, Swecker WS Jr, Jensen RV, and Ponder MA. Characterization of the fecal bacteria communities of forage-fed horses by pyrosequencing of 16S rRNA V4 gene amplicons. *FEMS Microbiol Lett*. 2012; 326(1): 62-68. DOI: <https://doi.org/10.1111/j.1574-6968.2011.02434.x>
55. Dougal K, de la Fuente G, Harris PA, Girdwood SE, Pinloche E, and Newbold JC. Identification of a core bacterial community within the large intestine of the horse. *PLoS ONE*. 2013; 8(10): e77660. DOI: <https://doi.org/10.1371/journal.pone.0077660>
56. World Health Organization / Food and Agriculture Organization of the United Nations (FAO/WHO). Guidelines for the evaluation of probiotics in food. 2002. Available at: https://4cau4jsaler1zgkq3wnmje1-wpengine.netdna-ssl.com/wp-content/uploads/2019/04/probiotic_guidelines.pdf
57. Oelschlaeger TA. Mechanisms of probiotic actions—A review. *Int J Med Microbiol*. 2010; 300(1): 57-62. DOI: <https://doi.org/10.1016/j.ijmm.2009.08.005>
58. Desrochers AM, Dolente BA, Roy MF, Boston R, and Carlisle S. Efficacy of *Saccharomyces boulardii* for treatment of horses with acute enterocolitis. *J Am Vet Med Assoc*. 2005; 227(6): 954-959. DOI: <https://doi.org/10.2460/javma.2005.227.954>
59. Boyle AG, Magdesian KG, Durando MM, Gallop R, and Sigdel S. *Saccharomyces boulardii* viability and efficacy in horses with antimicrobial-induced diarrhoea. *Vet Rec*. 2013; 172(5): 128. DOI: <https://doi.org/10.1136/vr.100833>
60. Weese JS, Anderson ME, Lowe A, Penno R, da Costa TM, Button L, et al. Screening of the equine intestinal microflora for potential probiotic organisms. *Equine Vet J*. 2004; 36: 351-355. DOI: <https://doi.org/10.2746/0425164044890616>
61. Schoster A, Staempfli HR, Abrahams M, Jalali M, Weese JS, and Guardabassi L. Effect of a probiotic on prevention of diarrhea and *Clostridium difficile* and *Clostridium perfringens* shedding in foals. *J Vet Intern Med*. 2015; 29(3): 925-931. DOI: <https://doi.org/10.1111/jvim.12584>
62. Strobel C, Gunther E, Romanowski K, Busing K, Urubschuro V, and Zeyner A. Effects of oral supplementation of probiotic strains of *Lactobacillus rhamnosus* and *Enterococcus faecium* on diarrhoea events in foals in their first weeks of life. *J Anim Physiol Anim Nutr*. 2018; 102(5): 1357-1365. DOI: <https://doi.org/10.1111/jpn.12923>
63. Yuyama T, Shigeki Yusa, Takai S, Tsubaki S, Kado Y, and Morotomi M. Evaluation of a host-specific *Lactobacillus* probiotic in neonatal foals. *J Appl Res Vet Med*. 2004; 2: 26-32. Available at: <https://bluegrassanimalproducts.com/wp-content/uploads/2018/03/foal-study.pdf>
64. Parraga ME, Spier SJ, Thurmond M, and Hirsh D. A clinical trial of probiotic administration for prevention of *Salmonella* shedding in the postoperative period in horses with colic. *J Vet Intern Med*. 1997; 11: 36-41. DOI: <https://doi.org/10.1111/j.1939-1676.1997.tb00071.x>
65. Kim LM, Morley PS, Traub-Dargatzis JL, Salman MD, and Gentry-Weeks C. Factors associated with *Salmonella* shedding among equine colic patients at a veterinary teaching hospital. *J Am Vet Med Assoc*. 2001; 218(5): 740-748. DOI: <https://doi.org/10.2460/javma.2001.218.740>
66. Patro JN, Ramachandran P, Barnaba T, Mammel MK, Lewis JL, and Elkins CA. Culture independent metagenomic surveillance of commercially available probiotics with high throughput next generation sequencing. *mSphere*. 2016; 1(2): e00057-16. DOI: <https://doi.org/10.1128/mSphere.00057-16>
67. Berreta A, Baumgardner RM, Kopper JJ. Evaluation of commercial veterinary probiotics containing enterococci for transmissible vancomycin resistance genes. *BMC Res Notes*. 2020; 13:275
68. Minalyan A, Gabrielyan L, Scott D, Jacobs J, and Pisegna JR. The gastric and intestinal microbiome: Role of proton pump inhibitors. *Curr Gastroenterol Rep*. 2017; 19(8): 42. DOI: <https://doi.org/10.1007/s11894-017-0577-6>
69. Modesto M, D'Aimmo MR, Stefanini I, et al. A novel strategy to select *Bifidobacterium* strains and prebiotics as natural growth promoters in newly weaned pigs. *Livest Sci*. 2009; 122:248-58.
70. Böhmer BM, Kramer W, Roth-Maier DA. Dietary probiotic supplementation and resulting effects on performance, health status and microbial characteristics of primiparous sows. *J Anim Physiol Anim Nutr*. 2006; 90:309-15.
71. Hong, H.A., Hong Duc, L. and Cutting, S., 2005. The use of bacterial spore formers as probiotics. *FEMS Microbiology Reviews* 29: 813-835.
72. Hassoun-Kheir N, Stabholz Y, Kreft JU, de la Cruz R, Romalde JL, Nesme J, et al. Comparison of antibiotic-resistant bacteria and antibiotic resistance genes abundance in hospital and community wastewater: A systematic review. *Sci Total Environ*. 2020; 743: 140804. DOI: <https://doi.org/10.1016/j.scitotenv.2020.140804>
73. Da Costa PM, Loureiro L, and Matos AJF. Transfer of multidrug-resistant bacteria between intermingled ecological niches: The interface between humans, animals and the environment. *Int J Environ Res Public Health*. 2013; 10(1): 297-294. DOI: <https://doi.org/10.3390/ijerph10010278>
74. Banerjee G and Ray AK. The advancement of probiotics research and its application in fish farming industries. *Res Vet Sci*. 2017; 115: 66-77. DOI: <https://doi.org/10.1016/j.rvsc.2017.01.016>
75. Olmos J, Acosta M, Mendoza G, and Pitones V. *Bacillus subtilis*, an ideal probiotic bacterium to shrimp and fish aquaculture that increase feed digestibility, prevent microbial diseases, and avoid water pollution. *Arch Microbiol*. 2020; 202(3): 427-435. DOI: <https://doi.org/10.1007/s00203-019-01757-2>
76. Balcázar JL, Rojas-Luna T, and Cunningham DP. Effect of the addition of four potential probiotic strains on the survival of Pacific white shrimp (*Litopenaeus vannamei*) following immersion challenge with *Vibrio parahaemolyticus*. *J Invertebr Pathol*. 2007; 96(2): 147-150. DOI: <https://doi.org/10.1016/j.jip.2007.04.008>
77. Giri SS, Sukumaran V, and Oviya M. Potential probiotic *Lactobacillus plantarum* VSG3 improves the growth, immunity, and disease resistance of tropical freshwater fish, *Labeo rohita*. *Fish Shellfish Immunol*. 2013; 34(2): 660-662. DOI: <https://doi.org/10.1016/j.fsi.2012.12.008>
78. Marques A, Thanh TH, Sorgeloos P, and Bossier P. Use of microalgae and bacteria to enhance protection of gnotobiotic *Artemia* against different pathogens. *Aquaculture*. 2006; 258(1-4): 116-126. DOI: <https://doi.org/10.1016/j.aquaculture.2006.04.021>
79. Supamattaya K, Kiriratnikom S, Boonyaratpalin M, and Borowitzka L. Effect of a *Dunaliella* extract on growth performance, health condition, immune response and disease resistance in black tiger shrimp (*Penaeus monodon*). *Aquac*. 2005; 248(1-4): 207-216. DOI:

- <https://doi.org/10.1016/j.aquaculture.2005.04.014>
80. Mo WY, Choi WM, Man KY, and Wong MH. Food waste-based pellets for feeding grass carp (*Ctenopharyngodon idellus*): Adding baker's yeast and enzymes to enhance growth and immunity. *Sci Total Environ*. 2020; 707: 134954. DOI: <https://doi.org/10.1016/j.scitotenv.2019.134954>
 81. Wu R, Shen J, Tian D, Yu J, He T, Yi J, et al. A potential alternative to traditional antibiotics in aquaculture: Yeast glycoprotein exhibits antimicrobial effect *in vivo* and *in vitro* on *Aeromonas caviae* isolated from *Carassius auratus gibelio*. *Vet Med Sci*. 2020; 6(3): 639-648. DOI: <https://doi.org/10.1002/vms3.253>
 82. Moriarty DJW. Control of luminous *Vibrio* species in penaeid aquaculture ponds. *Aquac*. 1998; 164(1-4): 351-358. DOI: [https://doi.org/10.1016/S0044-8486\(98\)00199-9](https://doi.org/10.1016/S0044-8486(98)00199-9)
 83. Hai NV. The use of probiotics in aquaculture. *J Appl Microbiol*. 2015; 119(4): 917-935. DOI: <https://doi.org/10.1111/jam.12886>
 84. Hai NV, Buller N, and Fotadar R. Effects of probiotics (*Pseudomonas synxantha* and *Pseudomonas aeruginosa*) on the growth, survival and immune parameters of juvenile western king prawns (*Penaeus latisulcatus* kishinouye, 1896). *Aquac Res*. 2009a; 40(5): 590-602. DOI: <https://doi.org/10.1111/j.1365-2109.2008.02135.x>
 85. Aly SM, Mohamed MF, and John G. Effect of probiotics on the survival, growth and challenge infection in *Tilapia nilotica* (*Oreochromis niloticus*). *Aquac Res*. 2008; 39(6): 647-656. DOI: <https://doi.org/10.1111/j.1365-2109.2008.01932.x>
 86. Van Doan H, Hoseinifar SH, Tapingkae W, Seel-audom M, Jaturasitha S, Dawood MAO, et al. Boosted growth performance, mucosal and serum immunity, and disease resistance Nile Tilapia (*Oreochromis niloticus*) Fingerlings Using Corn-cob-Derived Xylooligosaccharide and *Lactobacillus plantarum* CR1T5. *Probiotics Antimicrob Proteins*. 2020; 12(2): 400-411. DOI: <https://doi.org/10.1007/s12602-019-09554-5>
 87. Waiyarnit P, Zoral MA, Saengtienchai A, Luengnaruemitchai A, Decamp O, Gorgoglione B, et al. Probiotics modulate tilapia resistance and immune response against tilapia lake virus infection. *Pathogens*. 2020; 9(11): 919. DOI: <https://doi.org/10.3390/pathogens9110919>
 88. Chythanya R, Karunasagar I, and Karunasagar I. Inhibition of shrimp pathogenic vibrios by a marine *Pseudomonas* I-2 strain. *Aquac*. 2002; 208(1-2): 1-10. DOI: [https://doi.org/10.1016/S0044-8486\(01\)00714-1](https://doi.org/10.1016/S0044-8486(01)00714-1)
 89. Castex M, Chim L, Pham D, Lemaire P, Wabete N, Nicolas JL, et al. Probiotic P. acidilactici application in shrimp *Litopenaeus stylirostris* culture subject to vibriosis in New Caledonia. *Aquac*. 2008; 275(1-4): 182-193. DOI: <https://doi.org/10.1016/j.aquaculture.2008.01.011>
 90. Wang R, Guo Z, Tang Y, Kuang J, Duan Y, Lin H, et al. Effects on development and microbial community of shrimp *Litopenaeus vannamei* larvae with probiotics treatment. *AMB Express*. 2020; 10: 10. DOI: <https://doi.org/10.1186/s13568-020-01041-3>
 91. Leyva-Madrigal KY, Luna-González A, Escobedo-Bonilla CM, Fierro-Coronado JA, and Maldonado-Mendoza IE. Screening for potential probiotic bacteria to reduce prevalence of WSSV and IHHNV in whiteleg shrimp (*Litopenaeus vannamei*) under experimental conditions. *Aquac*. 2011; 322-323: 16-22. DOI: <https://doi.org/10.1016/j.aquaculture.2011.09.03>
 92. Gunasundari V, Ajith Kumar TT, Ghosh S, and Kumaresan S. An *ex vivo* tool to evaluate the brewer's yeast *Saccharomyces cerevisiae* in clownfish aquaculture with special reference to *Amphiprion percula* (Lacepede, 1802). *Turkish J Fish Aquat Sci*. 2013; 13: 389-395. Available at: <https://www.trjfas.org/uploads/pdf/192.pdf>
 93. Li P and Gatlin DM. Evaluation of the prebiotic GroBiotic®-A and brewers yeast as dietary supplements for sub-adult hybrid striped bass (*Morone chrysops* x *M. saxatilis*) challenged *in situ* with *Mycobacterium marinum*. *Aquac*. 2005; 248(1-4): 197-205. DOI: <https://doi.org/10.1016/j.aquaculture.2005.03.005>
 94. Li P and Gatlin DM. Dietary brewers yeast and the prebiotic GroBiotic™ AE influence growth performance, immune responses and resistance of hybrid striped bass (*Morone chrysops* x *M. saxatilis*) to *Streptococcus iniae* infection. *Aquaculture*. 2004; 231(1-4): 445-456. DOI: <https://doi.org/10.1016/j.aquaculture.2003.08.021>
 95. Taoka Y, Maeda H, Jo JY, Jeon MJ, Bai SC, Lee WJ, et al. Growth, stress tolerance and non-specific immune response of Japanese founder *Paralichthys olivaceus* to probiotics in a closed recirculating system. *Fish Sci*. 2006; 72(2): 310-321. DOI: <https://doi.org/10.1111/j.1444-2906.2006.01152.x>
 96. Midhun SJ, Neethu S, Arun D, Vysakh A, Divya L, Radhakrishna EK, et al. Dietary supplementation of *Bacillus licheniformis* HGA8B improves growth parameters, enzymatic profile and gene expression of *Oreochromis niloticus*. *Aquac*. 2019; 503: 289-296. DOI: <https://doi.org/10.1016/j.aquaculture.2019.02.064>
 97. Kuebutornye FKA, Abarike ED, Lu Y, Hlordzi V, Sakyi ME, Afriyie G, et al. Mechanisms and the role of probiotic *Bacillus* in mitigating fish pathogens in aquaculture. *Fish Physiol Biochem*. 2020; 46: 819-841. DOI: <https://doi.org/10.1007/s10695-019-00754-y>
 98. Arellano-Carbajal F and Olmos-Soto J. Thermostable α -1,4- and α -1,6-glucosidase enzymes from *Bacillus* sp. isolated from a marine environment. *World J Microbiol Biotechnol*. 2002; 18: 791-795. Available at: <https://link.springer.com/article/10.1023/A:1020433210432>
 99. Leonel Ochoa-Solano J and Olmos-Soto J. The functional property of *Bacillus* for shrimp feeds. *Food Microbiol*. 2006; 23(6): 519-525. DOI: <https://doi.org/10.1016/j.fm.2005.10.004>
 100. Chen B, Peng M, Tong W, Zhang Q, and Song, Z. The Quorum Quenching Bacterium *Bacillus licheniformis* T-1 protects Zebrafish against *Aeromonas hydrophila* infection. *Probiotics Antimicrob Proteins*. 2020a; 12: 160-171. Available at: <https://link.springer.com/article/10.1007/s12602-018-9495-7>
 101. Chen YY, Wang YL, Wang WK, Zhang ZW, Si XM, Cao ZJ, et al. Beneficial effect of *Rhodopseudomonas palustris* on *in vitro* rumen digestion and fermentation. *Benef Microbes*. 2020b; 11(1): 91-99. DOI: <https://doi.org/10.3920/BM2019.0044>
 102. Kewcharoen W and Srisapoom P. Probiotic effects of *Bacillus* spp. from Pacific white shrimp (*Litopenaeus vannamei*) on water quality and shrimp growth, immune responses, and resistance to *Vibrio parahaemolyticus* (AHPND strains). *Fish Shellfish Immunol*. 2019; 94: 175-189. DOI: <https://doi.org/10.1016/j.fsi.2019.09.013>
 103. Dalmin G, Kathiresan K, and Purushothaman A. Effect of probiotics on bacterial population and health status of shrimp in culture pond ecosystem. *Indian J Exp Bio*. 2001; 39(9): 939-942. Available at: <https://pubmed.ncbi.nlm.nih.gov/11831382/>
 104. Wang Y and He Z. Effect of probiotics on alkaline phosphatase activity and nutrient level in sediment of shrimp, *Penaeus vannamei*, ponds. *Aquac*. 2009; 287(1-2): 94-97. DOI: <https://doi.org/10.1016/j.aquaculture.2008.10.022>
 105. Wang YB, Xu ZR, and Xia MS. The effectiveness of commercial probiotics in northern white shrimp *Penaeus vannamei* ponds. *Fish Sci*. 2005; 71: 1036-1041. DOI: <https://doi.org/10.1111/j.1444-2906.2005.01061.x>
 106. Babazadeh D, Vahdatpour T, Nikpiran H, Jafarholipour MA and Vahdatpour S. Effects of probiotic, prebiotic and synbiotic intake on blood enzymes and performance of Japanese quails (*Coturnix japonica*). *Indian J of Anim Sci*. 2011;81(8): 870-874.
 107. Nikpiran H, Vahdatpour T, Babazadeh D, Tabatabaei SM, and Vahdatpour S. Effects of Functional Feed Additives on Growth Influenced Hormones and Performance of Japanese Quails (*Coturnix japonica*). *Greener J of Bio Sci*. 2014;4 (2): 039-044. Available at: <https://www.scinapse.io/papers/2323575296#fullText>
 108. Babazadeh D, and Asasi K. Effects of *in ovo* synbiotic injection on growth performance, intestinal bacterial load and antibody titres in broiler chickens vaccinated against infectious bursal disease. *Bulgarian Journal of Veterinary Medicine*, 2021, 24, No 4, 520-532. DOI: <https://doi.org/10.15547/bjvm.2298>
 109. Saeed M, Babazadeh D, Naveed M, Alagawany M, E Abd El-Hack M, Arain MA, Tiwari R, Sachan S, Karthik K, Dhama K, Elnesrki SS and Chao S. *In ovo* delivery of various biological supplements, vaccines and drugs in poultry: current knowledge. *J Sci Food Agric* 2019; 99: 3727-3739. DOI: <https://doi.org/10.1002/jsfa.9593>
 110. Nikpiran H, Vahdatpour T, Babazadeh D, and Vahdatpour S. Effects of *Saccharomyces cerevisiae*, Thpax and their combination on blood enzymes and performance of Japanese quails (*Coturnix japonica*). *Journal of Animal and Plant Sciences*, 2013;23: 369-375. Available at: <https://www.semanticscholar.org/paper/Effects-of-Saccharomyces-cerevisiae%2C-Thpax-and-on-Nikpiran-Vahdatpour/becaf180aa466a89056b96812d10e6c35a2abbf0>
 111. Vahdatpour T, and Babazadeh D. The effects of Kefir rich in probiotic administration on serum enzymes and performance in male Japanese quails. *Journal of Animal & Plant Sciences*, 2016;26(1): 34-39. Available at: <http://thejaps.org.pk/docs/v-26-01/05.pdf>
 112. Saeed M, Babazadeh D, Arain MA, Naveed M, Shah QA, Kamboh AA, Moshaveri A, Modarresi-Ghazani F, Hejazi V, and Chao S. The use of chicoric acid from *Echinacea purpurea* as a feed additive in

- poultry nutrition. *World's Poultry Science Journal*. 2017;74(1): 69-78. DOI: <https://doi.org/10.1017/S0043933917001027>
113. Lee S; Lee J, Jin YI, Jeong JC, Chang YH, Lee Y, Jeong Y, and Kim M. Probiotic characteristics of *Bacillus* strains isolated from Korean traditional soy sauce. *LWT Food Sci. Technol*. 2017; 79: 518-524. DIO: <https://doi.org/10.1016/j.lwt.2016.08.040>
114. Vankerckhoven V, Huys G, Vancanneyt M, Vael C, Klare I, Romond M, Entenza JM, Moreillon P, Wind RD, Knol J, et al. Biosafety assessment of probiotics used for human consumption: Recommendations from the EU-PROSAFE project. *Trends Food Sci. Technol*. 2008; 19: 102-114. DOI: <https://doi.org/10.1016/j.tifs.2007.07.013>